THE STRUCTURE OF DETOXIN D

A SELECTIVE ANTAGONIST OF BLASTICIDIN S

Katsumi Kakinuma, Noboru Ötake and Hiroshi Yonehara

Institute of Applied Microbiology, The University of Tokyo, Tokyo

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Detoxin D_1 , one of the most active principle of detoxin complex produced by <u>Streptomyces</u> <u>caespitosus</u> var. <u>detoxicus</u>, is a selective antagonist of blasticidin S and shows several interesting biological activities to counteract the toxicity of the antibiotic against plant and animal cells.^{1),2)} This communication concerns with the structural elucidation of detoxin D_1 .

Detoxin $D_1 \underline{I}$, $C_{28}H_{41}N_3O_8$, mp. 156 - 158°C, $[\alpha]_p^{25}$ -16° (C 1, MeOH), is an amphoteric compound with pKa 4.0 and 8.0 and shows positive ninhydrin reaction. \underline{I} is a kind of peptide $(\bigvee_{max}^{wajel} 3400, 2750, 1740, 1650, 1600 \text{ cm}^{-1})$ exhibiting the signals of one phenyl nucleus $(\int_{0}^{COCL_3} 7.22)$, one acetyl (§ 2.00) and four methyl groups (near § 1.0) in the NMR spectrum.

Detoxin D_1 acetate methyl ester <u>II</u>, $C_{31}H_{45}N_3O_9$ (M⁺: m/e=603) was obtained by acetylation of <u>I</u> with acetic anhydride in pyridine followed by esterification with diazomethane.

Acid hydrolysis of \underline{I} with 5.7N HCl at 110° C for 16 hrs provided each one mole of L-valine and L-phenylalanine accompanying an unknown amino acid designated detoxinine which was coloured to yellow with ninhydrin. The N-terminus was determined to the amino group of L-valine by DNP and diazotization methods.

Fig. 1

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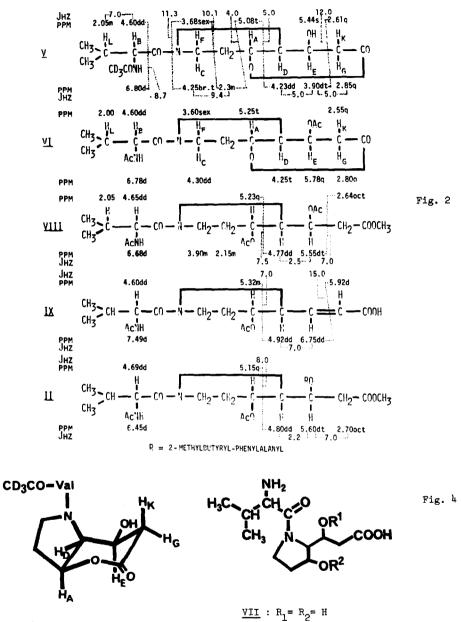


Fig. 3

 $\underbrace{\mathbf{r}_1}_{1=1} \cdot \mathbf{r}_1 = \mathbf{r}_2 = \mathbf{n}$ $\underbrace{\mathbf{Ia}}_{1=1} \cdot \mathbf{R}_1 = \mathbf{Ac}, \mathbf{R}_2 = 2 - \mathbf{methylbutyrylphenylalanyl}$ $\underbrace{\mathbf{Ib}}_{1=1} \cdot \mathbf{R}_2 = \mathbf{Ac}, \mathbf{R}_1 = 2 - \mathbf{methylbutyrylphenylalanyl}$

No. 25

Alkaline hydrolysis of <u>I</u> with 0.1N NaOH at room temperature for a week, followed by ethereal extraction at acidic condition, gave crystalline needles <u>III</u>, $C_{14}H_{19}NO_3$ (M⁺: m/e=249) mp. 122-123°C, γ_{max}^{nujel} 3300, 2800, 1740, 1650 cm⁻¹. L-Phenylalanine and (+)-S-2-methylbutyric acid, $[\alpha]_p^{22}$ + 19.2° (C 1.2, MeOH)³, were obtained by acid hydrolysis of <u>III</u>. These evidences together with the NMR and mass spectral data are consistent with the structure of (+)-2-methylbutyryl-L-phenylalanine for <u>III</u>.

Alkaline hydrolysis of <u>I</u> with 1N NaOH followed by resin chromatography on Dowex 50W x 2 [H⁺] gave valul-detoxininolactone <u>IV</u>, which was acylated with CD_3COCl in pyridine to crystalline N-deuteroacetate <u>V</u>, $C_{14}H_{19}N_2O_5D_3$ (M⁺: found; m/e=301.1744, calcd.; 301.1715), mp. 174 - 175°C, γ_{max}^{KBr} 1730 cm⁻¹ and with acetic anhydride in pyridine to diacetate <u>VI</u>, $C_{16}H_{24}N_2O_6$ (M⁺: m/e=340)

The results of NMR and spin decoupling experiments of <u>V</u> and <u>VI</u> together with the IR and mass spectral data provided the evidences of their structures as formula <u>V</u> and <u>VI</u> (Fig. 2 and 3).

On the other hand, the resin chromatography on Dowex 50W x 2 of the alkaline hydrolysate of I after neutralization with dilute hydrochloric acid provided amorphous powder of valyldetoxinine <u>VII</u>, $C_{12}H_{22}N_2O_5$. Unlike the lactone <u>IV</u>, <u>VII</u> has an amphoteric nature with a free carboxyl group ($\sqrt[N_{max}]_{wax}^{nujel}$ 2700, 1730 cm⁻¹), which was converted to the corresponding triacetate methyl ester <u>VIII</u>, $C_{19}H_{30}N_2O_8$ (M⁺: m/e=414), on acetylation followed by esterification. The NMR spectrum of <u>VIII</u> is also shown in Fig. 2 and is very similar to that of <u>II</u> except the signals of 2-methylbutyryl-phenylalanyl moiety. Accordingly, the structure of <u>VIII</u> was established (Fig. 4).

These accumulated informations lead to the structure <u>Ia</u> or <u>Tb</u> for detoxin D_1 , and the binding position of <u>III</u> remains to be settled unequivocally. Several attempts in effecting partial hydrolysis of <u>I</u> or transesterification of bulky group <u>III</u> were unsuccessful. Since the β -acyloxycarboxylic group is present in the structure of <u>Ia</u> or <u>Ib</u>, β -elimination provoked by the nucleophiles to form an α , β -unsaturated carboxylic acid would be expected. Fortunately, treatment of <u>I</u> with acetic anhydride at 70°C for 1 hr yielded the α , β -unsaturated acid Nacetate <u>IX</u> $C_{16}H_{24}N_2O_6$ (M⁺: m/e=340), λ_{max}^{E+OH} 210 nm (ε 14,000), \S 5.92 (1H,d), 6.75 (1H,dd).

This result demonstrated that the bulky group <u>III</u> was removed by the β -elimination and consequently, the structure of detoxin D₁ is established as depicted in Fig. 1.

The absolute configuration of \underline{I} was determined as follows. There are six asymmetric carbons in \underline{I} and three of their configurations have been determined already. Remaining three are present in detoxinine moiety. Their absolute configurations were determined by the application of Klyne's lactone sector rule⁴ to \underline{V} .

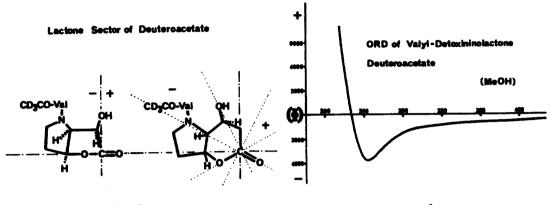


Fig. 5

Fig. 6

The relative configuration of \underline{V} was elucidated by its NMR coupling constant. The ring juncture of this lactone is <u>cis</u> $(J_{H_A}, H_p = 5_{HZ})$ and it has a chair conformation. The lactone sectors are shown in Fig. 5. The ORD of \underline{V} in MeOH showed clear negative Cotton effect at 243 nm as was expected from the sectors (Fig. 6). The coupling constants between H_A and H_D in <u>II</u> and <u>VIII</u> are 7.5 _{HZ} and 8.0_{HZ} in CDCl₃, respectively. The theoretical values calculated from the models are 7.7_{HZ} for <u>cis</u> and 4.9_{HZ} for <u>trans</u> configuration. Therefore, it is apparent that H_A and H_D in <u>II</u> are located in <u>cis</u> configuration since the inversion of configuration on the carbon adjacent to lactone oxygen did not take place.

On the basis of the evidences described above, the absolute structure of detoxin D_1 was established conclusively as <u>I</u> which is a kind of depsipeptide containing a new amino acid with pyrrolidine nucleus.

References

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